

## Brief Communication

# Chromosome-scale reference genome provides insights into the genetic origin and grafting-mediated stress tolerance of *Malus prunifolia*

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Grafting, an ancient horticultural cultivation technique, is essential for commercial apple (*Malus × domestica* Borkh.) production. *M. prunifolia* ‘Fupingqiuzi’ is used as a common apple rootstock due to its known tolerances to abiotic and biotic stresses. However, the genetic origin and evolutionary history of *M. prunifolia* are largely unknown, as are the molecular bases underlying its innate stress tolerance characteristics and ability to graft-transmit stress tolerance to the scion.

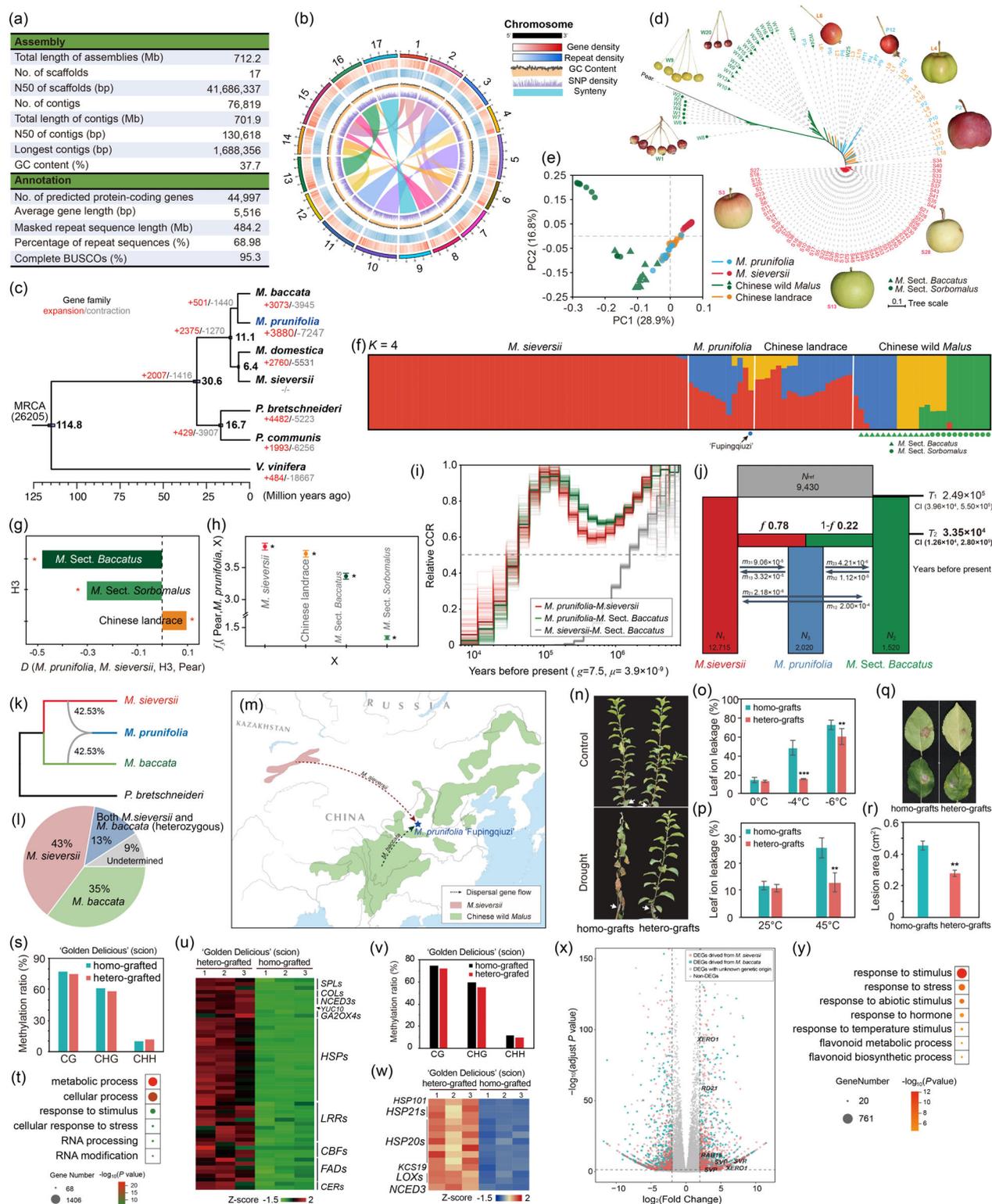
Here, we present a high-quality chromosome-scale genome sequence of *M. prunifolia* ‘Fupingqiuzi’ ( $2n = 2x = 34$ ) assembled using a combined strategy of Illumina short reads, PacBio long reads, and Hi-C data (Figure S1, Tables S1–S3). The total length of the assembly was 712 Mb, with a scaffold N50 of 41.6 Mb (Figure 1a). Approximately 98.6% of the contigs were anchored into 17 pseudochromosomes (Figure 1b). Over 91.36% of non-redundant and single-mapped mate-pair reads were concordant pairs, indicating that Hi-C anchoring has high accuracy. BUSCO revealed a completeness rate of 95.3% (Table S4). A total of 44 997 protein-coding genes were predicted, with an average gene length of 5516 bp. In addition, 1785 non-coding RNA genes and 484 Mb of repetitive sequences were detected in the ‘Fupingqiuzi’ genome (Figure 1a, Tables S5–S6).

Gene family analysis identified 3880 expanded gene families in *M. prunifolia* (Figure 1c), which were enriched mainly in response to stress, bacterium, abscisic acid (ABA), immune response, and

plant–pathogen interaction (Table S7, Data S1), with 56 homologs of *HSP* and 69 homologs of *LRR* were expanded significantly (Table S8).

To explore the genetic origin of the *M. prunifolia*, we sequenced 79 *Malus* accessions (Chen *et al.*, 2021) and downloaded 34 *Malus* resequencing data sets (Duan *et al.*, 2017), including 12 *M. prunifolia*, 58 *M. sieversii*, 18 Chinese landrace, and 25 Chinese wild *Malus* accessions (Table S9). In total, 931.35 Gb of whole-genome sequencing data were obtained, and 14 073 781 SNPs were identified (Figure 1b). Phylogenetic and principal component analysis (PCA) showed that *M. prunifolia* and Chinese landrace accessions clustered together (Figure 1d–e), demonstrating that their genetic structures are very similar and *M. prunifolia* is paraphyly. Genetic structure of *M. prunifolia* accessions exhibited an admixture pattern, compared with *M. sieversii* and *M. Sect. Baccatus* (Figure 1f, Figures S2–S4). *D*-statistics analysis of *M. prunifolia* and the other groups showed that *M. Sect. Baccatus* shared more alleles with *M. prunifolia* (Figure 1g, Table S10). Furthermore,  $f_3$  statistics illustrated that the *M. prunifolia* had a closer genetic affinity with *M. sieversii* (Figure 1h). These results indicate that *M. prunifolia* has a hybrid origin between *M. sieversii* and *M. Sect. Baccatus*.

To further confirm the hybrid origin of *M. prunifolia*, we performed inference of demographic history. The divergence times of *M. prunifolia* versus *M. sieversii* and *M. prunifolia* versus *M. Sect. Baccatus* were  $-49\,614$  (95% confidence interval: 37 589–57 888) and  $-49\,746$  (95% CI: 43 526–56 967) years before present, respectively, as estimated by the Multiple Sequentially Markovian Coalescent 2 (MSMC2) (Figure 1i). The two divergence times were almost identical, implying that *M. prunifolia* accessions have hybrid origin. Moreover, the simulation of diffusion approximations for demographic inference ( $\partial a \partial i$ ) showed that the hybrid origin model was the best-fitting model (Figure S5, Table S11). The estimated hybrid parameter ( $f$ ) indicated that  $\sim 78\%$  of the nuclear genome of the initial *M. prunifolia* came from *M. sieversii*, and  $\sim 22\%$  from *M. Sect. Baccatus*, consistent with genetic admixture. Speciation time of



**Figure 1** (a, b) Genome assembly, annotation (a), and features (b) of *M. prunifolia* 'Fupingqizui'. (c) Gene family analysis. (d-f) Phylogenetic tree (d), PCA (e), and admixture analysis (f) of all 113 *Malus* accessions. (g, h)  $D$  and  $f_3$  statistics analyses. Error bars represent SE. (i) Inferred relative cross-coalescence rate among *Malus* populations. (j) Simulation results of the best  $\partial a \partial i$  model. (k) ML-bootstrap network. (l, m) Genetic contribution of the two main ancestors to 'Fupingqizui'. (n) Drought, (o) Freezing, (p) Thermotolerance, and (q, r) Disease resistance of homo- and hetero-grafts. Data are means  $\pm$  SD. (s, v) Relative proportions of mCs (methylated cytosines) in homo- and hetero-grafted 'Golden Delicious' under control (s) and drought conditions (v). (t) GO enrichment analysis of DMR-related genes between homo- and hetero-grafted 'Golden Delicious'. (u, w) Expression of DEGs between homo- and hetero-grafted 'Golden Delicious' under (u) control and drought conditions (w). (x, y) Genetic origins (x) and functional enrichment (y) of DEGs in hetero-grafted 'Fupingqizui' in response to drought stress. Three biological replicates were prepared for each condition in (s-y).

*M. prunifolia* was dated to ~33 500 years before present, close to the results by MSMC2 (Figure 1j, Figure S6, and Table S12).

Phylogenetic networks from the 2756 orthologous single-copy gene tree topologies showed reticulately evolutionary relationships among 'Fupingqiuzi', *M. sieversii*, and *M. baccata* genomes (Figure 1k), indicating a hybrid origin for 'Fupingqiuzi'. A window-based genetic distance approach also showed that approximately 43% of the *M. prunifolia* genome was derived from *M. sieversii* and 35% derived from *M. baccata* (Figure 1l), further supporting the hybrid origin of 'Fupingqiuzi' from ancient admixture of *M. sieversii* and *M. baccata* (Figure 1m, Figure S7, Table S13).

To investigate the influence of 'Fupingqiuzi' (as rootstocks) on stress resistance of scion, we generated homo-grafts (*M. × domestica* 'Golden Delicious' grafted onto itself) and hetero-grafts ('Golden Delicious' grafted onto 'Fupingqiuzi'). We found that compared with homo-grafts, hetero-grafts were more tolerant to drought, extreme temperatures, and pathogen infection (Figure 1n-r).

To reveal the molecular basis of grafting-mediated stress tolerance, we analysed the genome-wide DNA methylation and gene expression of homo- and hetero-grafted 'Golden Delicious'. Compared with homo-grafted 'Golden Delicious', CG and CHG methylation (H = adenine, thymine, or cytosine) were higher, whereas CHH was lower in the hetero-grafted 'Golden Delicious' but not to a significant level (Figure 1s, Figure S8). Differentially methylated regions (DMRs) analysis between hetero- and homo-grafted 'Golden Delicious' identified 4002 DMRs and 3188 DMR-related genes, with their encoded proteins significantly enriched in terms of 'cellular response to stress' (Figure 1t, Data S2-S3). Among the DMR-related genes, 652 genes were involved in the environmental adaptation, and some genes were related to flowering and ABA biosynthesis (Data S2). In addition, compared with homo-grafts, hetero-grafts exhibited 2750 differentially expressed genes (DEGs) in scions. Among these DEGs, we identified flowering-related genes including homologs of *SPL4*, *SPL13*, *CONSTANS-LIKE 2 (COL2)*, and genes related to the biosynthesis of hormones including homologs of *NCED3*, *YUC10*, *GA2OX6*, and *GA2OX1* (Data S4). In addition, the up-regulated genes contained stress-responsive genes, including homologs of *HSPs*, *LRRs*, *CBFs*, *FADs*, and cuticular wax biosynthesis genes (Figure 1u). DNA methylation modifies chromatin structure and thereby affects the transcriptional regulation of genes (Zhang *et al.*, 2018). Among the DMR-associated DEGs, genes related to flowering and stress adaptation in hetero-grafted 'Golden Delicious', including homologs of *FAD4*, *SPL5*, *LRR*, and *DJC76*.

To further investigate the response of grafting combination to stress tolerance, we performed drought stress treatment on homo- and hetero-grafted 'Golden Delicious'. Genome-wide methylation levels were reduced in all CG, CHG, and CHH sequence contexts in hetero-grafted 'Golden Delicious' relative to homo-grafted 'Golden Delicious' under drought conditions (Figure 1v, Figure S9). We identified a total of 2512 DMR-related genes, which were mainly related to stimulus (Figure S9, Data S5-S6). Moreover, there are 497 DEGs in hetero-grafted 'Golden Delicious' compared with homo-grafted 'Golden Delicious',

which are strongly associated with abiotic stresses (Figure S9, Data S7-S8). Notably, we found that genes related to molecular chaperones, biosynthesis of cuticular wax and ABA, and lipoxygenase were highly expressed in hetero-grafted 'Golden Delicious', these included homologs of *HSP*, *KCS19*, *LOX*, and *NCED3* (Figure 1w).

We also sequenced the transcriptomes of homo- and hetero-grafted 'Fupingqiuzi' (rootstocks) under drought treatment. A total of 3102 DEGs were significantly enriched to the abiotic stresses (Figure 1x-y, Data S9 and S10). More importantly, 1271 and 1147 DEGs were derived from *M. sieversii* and *M. baccata*, respectively (Figure 1x). Notably, DEGs derived from *M. sieversii* including homologs of *XERO1* and *RAB18* were up-regulated whereas expression of genes derived from *M. baccata* including homologs of *RD21* and *SVP* was increased in 'Fupingqiuzi' under drought stress, suggesting the different gene contributions of *M. sieversii* and *M. baccata* to the 'Fupingqiuzi' (Figure 1x, Data S11).

In summary, these findings provide insights into the genetic origin of Chinese-originated crabapples and lay the foundation for a better understanding grafting-mediated stress tolerance.

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## Conflict of interest

No conflict of interest was declared.

## Author contributions

Q.G., Q.X., and F.M. conceived the study. Other authors carried out experiments and analyses. Z.L. and J.H. wrote the manuscript.

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## Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

**Figure S1-S11** Supplementary Figures.

**Data S1-S11** Supplementary Datasets.

**Methods S1** Supplementary Methods.

**Table S1-S13** Supplementary Tables.